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DescriptionField of the Invention

5 This invention relates to a method of separating and purifying a polypeptide from a complex aqueous mixture. More specifically, the invention relates to such a method in which the polypeptide-containing mixture undergoes a two-step chromatographic adsorption procedure, which procedure comprises an antibody purification step and an affinity region purification step.

10 Background of the Invention

In recent years, the scientific and medical communities have given increased attention to various polypeptides useful as therapeutic agents and to methods of isolating such polypeptides from the complex source materials in which they are present. An example of such a polypeptide is a blood factor obtained from plasma known as the antihemophilic factor. This blood factor is also identified as Factor VIII
15 procoagulant activity protein (Factor VIII:C). This protein acts to correct the clotting defect in individuals with hemophilia A. It exists in plasma complexed with another protein known as Factor VIII-related protein or Factor VIII:RP. Other designations for Factor VIII:RP are Factor VIII:R:Ag and von Willebrand factor. Because of Factor VIII:C's therapeutic value as a coagulant, it has been regarded as desirable to purify Factor VIII and to isolate Factor VIII:C from Factor VIII:RP. Various procedures have been suggested for the isolation and purification of Factor VIII:C and other polypeptides of therapeutic value. These methods have been generally based on the techniques of immunoaffinity, affinity or ion exchange chromatography. For example, the method in a recent patent for the ultrapurification of Factor VIII:C, Zimmerman & Fulcher, U.S. Reissue Patent 32,011, employs such a two-step procedure of affinity and ion-exchange chromatography.
20 Essentially, a Factor-VIII preparation is passed through a column containing agarose beads coupled with mouse monoclonal antibodies directed to Factor VIII:RP. The Factor VIII:C, which is complexed to the von Willebrand factor, is adsorbed onto the matrix while uncomplexed Factor VIII:C moieties and contaminants pass through the column as unbound material. The Factor VIII:C is removed from the bound von Willebrand: antibody complex with a high salt solution containing calcium. The Factor VIII:C solution is desalted and finally adsorbed onto an ion-exchange column, more specifically, agarose beads coupled with positively charged aminohexyl groups. The Factor VIII:C is desorbed from the column with a high salt solution.
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Although this method is suitable for use in the ultrapurification of a polypeptide, it lacks several important features which would improve the therapeutic safety of the product and facilitate its large-scale production. One distinctive feature of the Zimmerman et al reissue patent is that the monoclonal antibodies
35 are directed to another polypeptide (von Willebrand factor) that is usually in excess and considered associated with the polypeptide of interest (Factor VIII:C). Depending upon the source material, it is possible that as much as 50% of the Factor VIII:C can be in a form not associated with von Willebrand factor. See Amphlett et al, U.S. Patent 4,508,709. The unassociated Factor VIII:C will not be bound by the monoclonal antibodies used in the immunoaffinity step described above, and will consequently be lost in the purification process. To date, there has been no proven advantage for having Factor VIII:C as a product in the uncomplexed form only. Evidence suggests that Factor VIII:C is protected longer from proteolysis by its association with the von Willebrand factor, an important feature when isolating the polypeptide from complex source materials. See Weiss et al, *J. Clin. Invest.* 60, 390-404, 1977. Hence, instead of being a disadvantage, the association of von Willebrand factor with Factor VIII:C could be beneficial insofar as it
40 may confer stability to Factor VIII:C during the purification steps and may extend the half-life of the polypeptide during its therapeutic administration.

Secondly, monoclonal antibodies covalently coupled to any matrix have a tendency to leach, or separate from their matrix and contaminate the final polypeptide-containing product. The patented procedure described above and in the prior art does not guard against the probability of nonhuman cell-derived
50 leached monoclonal antibodies from the immunoaffinity step accompanying and reassociating with the Factor VIII:C during the second ion-exchange step. The high ionic strength buffer used in the immunoaffinity procedure to elute Factor VIII:C is reduced to low ionic strength, which could allow monoclonal antibodies removed from the immunoaffinity column either to rebind to Factor VIII:C or to bind and desorb from the ion-exchange matrix along with the Factor VIII:C.

55 Thirdly, the desalting process required for the polypeptide-containing solution prior to loading onto the ion exchange column is usually accomplished by large volume dilution, dialysis, or ultra-filtration molecular washing. These methods are not only cumbersome for large-scale production volumes, but inevitably lead to loss of product.

Finally, the aqueous source materials in which the polypeptides of interest are found often are contaminated with one or more viruses. There are techniques for inactivating viruses in polypeptide mixtures, but attempts to combine such techniques with known polypeptide purification processes have produced methods with a multiplicity of steps unsuitable for large-volume production. The methods have also frequently been only partially successful in purifying the polypeptide. For example, in the prior art, a number of viral-inactivating agents have been shown effective for inactivating viruses. The agents have, however, been either denaturing or difficult to separate from the polypeptide of interest, and have required a special treatment or separation step. Other conventional methods for treating polypeptide-containing preparations for potential viral contamination, such as heat or irradiation, have resulted in either significant denaturation of the polypeptide of interest and/or insufficient inactivation of viruses.

The use of viral inactivating agents described herein can inactivate viruses without adversely affecting the biological activity of the polypeptide of interest. Treatment of the aqueous source materials with the viral inactivating agents, accompanied by the other procedures of this invention, produces a final product substantially free of viruses as well as viral inactivating agents.

It is a principal object of this invention to provide a purification process for polypeptides particularly adapted to large-scale purification of a polypeptide with a low level of polypeptide denaturation. It is another object of the invention to provide a polypeptide purification process which is also effective in reducing antibody and viral contamination of the purified product. One additional object of this invention is the development of a process for purifying a polypeptide, free of contaminating substances such as other proteins, viruses, and treating agents used in the purification steps.

Summary of the Invention

In accordance with the present invention, there is disclosed a process for the isolation and purification of Factor VIII:C from a complex aqueous mixture. As a result of the process, the Factor VIII:C is substantially free of other contaminating proteins, and is further highly purified by comparison to its purity in the starting aqueous mixture. One preferred embodiment of this process is the inclusion in the process of a technique for reducing the level of pathogenic substances such as viruses. The method comprises purifying Factor VIII:C in a mixture of polypeptides and other constituents comprising subjecting the Factor VIII:C in said mixture to a multiple-stage purification process:

- (a) immobilizing an antibody, which binds by hydrophobic attraction to the Factor VIII:C to be purified, to a substrate before or after said Factor VIII:C is added to said antibody, thereby immobilizing said Factor VIII:C in an immunoaffinity matrix;
- (b) eluting the Factor VIII:C from the immobilized antibody by treating the Factor VIII:C immunoaffinity matrix with a desorbing substance to desorb the polypeptide from said matrix;
- (c) passing the Factor VIII:C to be purified through a second region capable of binding to the Factor VIII:C, thereby binding the Factor VIII:C through a hydrophilic attraction to the second region while allowing contaminants to pass through the second region, the Factor VIII:C-desorbing substance mixture being passed from the matrix of step (b) to the second region of step (c) without further modification of the said mixture; and
- (d) eluting the purified Factor VIII:C from the second region.

A preferred embodiment comprises incorporating into the above process a virus-inactivating agent comprising an organic solvent and a detergent, followed by removal of said solvent and detergent. This is advantageously done by purifying Factor VIII:C in a complex aqueous mixture contaminated with lipid-enveloped viruses by:

- (a) mixing the complex aqueous mixture with a virus-inactivating agent containing an organic lipid-dissolving or disrupting solvent and a detergent at any stage in the process;
- (b) passing the complex aqueous mixture through an immunoaffinity matrix containing active immobilized monoclonal antibody specific to the Factor VIII:C and which binds by hydrophobic attraction thereto, thereby causing the Factor VIII:C to adsorb to the immunoaffinity matrix;
- (c) washing the immunoaffinity matrix with a buffer to remove at least a portion of contaminants from the complex aqueous mixture, while leaving the Factor VIII:C adsorbed to the immunoaffinity matrix;
- (d) eluting the Factor VIII:C from the immunoaffinity matrix with an eluting substance, thereby forming a Factor VIII:C eluting substance mixture to be purified;
- (e) passing the said mixture to be purified through a second region which binds the Factor VIII:C by a hydrophilic attraction, thereby causing the polypeptide to bind thereto, while allowing any leached monoclonal antibodies, detergents and other contaminants to pass through the second region, wherein the said mixture to be purified is passed from the matrix of step (c) to the region of step (e) without

further modification of the mixture; and

(f) eluting the second region with an eluting solution to desorb the polypeptide to form a solution substantially free of contaminants.

5 Detailed Description of the Invention

The present invention is directed to a method of isolating and purifying Factor VIII:C from a complex aqueous mixture. Hereinafter, the terms "polypeptide of interest" and "polypeptide to be purified" refer to Factor VIII:C. By the process of this invention, Factor VIII:C can be isolated from complex mixtures such as blood plasma, plasma fractions, commercial concentrates, and tissue culture media, including such media containing synthetically produced polypeptides as well as naturally occurring polypeptides. The process of the invention produces Factor VIII:C substantially free of contaminants which were present in the mixture before processing as well as those added to the mixture during the processing steps disclosed below. Examples of contaminants include pathogenic viruses, both lipid-enveloped viruses and non-enveloped viruses, pyrogens, leached antibodies, organic solvents, detergents, desorbing agents and other proteins present in the complex mixture from which the polypeptide of interest is isolated.

In accordance with one embodiment of the process of this invention, a complex aqueous mixture containing Factor VIII:C is added to an antibody which binds to the Factor VIII:C to be purified. At the time the antibody binds to the polypeptide (Factor VIII:C), the antibody can be unattached to any other matrix, or it can be, prior to its binding to the polypeptide, bound to a matrix which immobilizes the antibody and fixes it within a predetermined region. A preferred type of region within which the antibody can be fixed is beaded resin in a chromatography column or radial flow cartridge. Whether or not the antibody is immobilized in a region before its binding to the polypeptide, it is important that the antibody be in an immobilized state at some point after the binding of antibody to polypeptide.

The antibody can be any antibody which is capable of being fixed to an immobilized matrix and able to bind to the polypeptide to be purified. Both polyclonal and monoclonal antibodies can be used, as well as mixtures of either type designed to bind with a number of immobilized substrates or polypeptides or both. Because of their specificity and ready availability in large quantities, monoclonal antibodies are preferred. The choice of antibody will depend upon the polypeptide and immobilized substrate chosen. The antibodies to be used herein with other preferred operating conditions are those which interact with the polypeptide through hydrophobic interaction.

A preferred method of this invention comprises binding the polypeptide to a monoclonal antibody specific to the polypeptide of interest after the antibody has itself been immobilized in an immunoaffinity chromatography column. The mixture of polypeptides is passed through the immunoaffinity chromatography column on which have been immobilized monoclonal antibodies specific to the polypeptide of interest. As the mixture passes through the column or cartridge, the polypeptide adsorbs to the immobilized monoclonal antibodies. The column or cartridge is washed with an aqueous buffer. The buffer removes a major portion of the virus-inactivating agent, as well as other contaminants, from the complex aqueous mixture, but leaves the Factor VIII:C adsorbed on the column or cartridge.

The polypeptide in the column is desorbed and eluted with a desorbing agent. Selection of desorbing agents is within the skill of the art. Preferred desorbing agents are low ionic strength, low polarity buffered solutions. Low ionic strength usually refers to solutions having an ionic strength, μ , less than 0.2. Examples of salt solutions with low ionic strength include 0.15 M sodium chloride, 1.0 mM calcium chloride, and 40 mM calcium chloride. Examples of nonpolar agents which provide low polarity solutions include ethylene glycol, dioxane, propylene glycol and polyethylene glycol. The buffer desorbs the polypeptide, thereby forming a low ionic strength, low polarity polypeptide solution.

The complex aqueous mixtures that provide a source of the Factor VIII:C often are contaminated by pathogenic viruses, especially lipid-enveloped viruses such as the hepatitis B virus, non-A and non-B hepatitis virus, and HIV (HTLV-III) virus. In the broadest applications of this invention, the immunoaffinity purification combined with the affinity purification, substantial quantities of virus are removed. When one of the preferred embodiments is employed by treating the polypeptide with an organic solvent and detergent, the levels of lipid-enveloped viruses are usually reduced below a detectable level. The operable components of this treatment are the organic solvents and the detergent. Organic solvents are those active in dissolving or disrupting the lipid-containing envelope surrounding many viruses and which do not denature the polypeptide to be purified. Examples include di- and trialkyl phosphates such as di-(n-propyl) phosphate and tri-(n-butyl) phosphate, ethers such as ethyl ether and propyl ether, esters such as amyl acetate, and alkylated and hydroxylated materials such as butylated hydroxyanisole (BHA) and butylated hydroxytoluene (BHT). Mixtures of the above or other organic solvents are also useful. Ethyl alcohol and ethyl ether have

been found to be particularly acceptable as have the alkyl phosphates.

Detergents useful herein can be chosen from any of the recognized groups of anionic, cationic and nonionic detergents. Examples include a number of sulfated alcohols and sodium acid salts such as sulfated oxyethylated alkylphenol (Triton W-30 and Triton X-100)), sodium dodecylbenzenesulfonate (Nacconol NR), sodium 2-sulfoethyl oleate (Igepon A), sodium cholate, sodium deoxycholate, sodium dodecylsulfonate, dodecyltrimethylbenzylammonium chloride (Triton K-60), oxyethylated amines (Ethomeen), N-dodecylaminoethanesulfonic acid, ethylene oxide-propylene oxide condensates (Pluronic copolymers), polyoxyethylated derivatives of esters (Tween 80 and Polysorbate 80), polyoxyethylene fatty alcohol ethers (Brij 35), Nonidet P-40 and Lubrox PX.

The amounts of organic solvent and detergent used in the practice of the preferred embodiments of this invention can vary, depending upon the aqueous mixture to be treated, and upon the solvent or detergent chosen. If ethers or alcohols or mixtures thereof are used, the amount can be from 1 to 50%, preferably from about 5 to 25% by weight of the aqueous mixture if that mixture is blood plasma or blood compositions. The alkyl phosphates are used in concentrations from 0.01 mg/ml of mixture treated to 1.0 g/ml, preferably between about 0.1 mg/ml and 10 mg/ml. The amount of detergent or wetting agent used is not critical. Its function is to improve the contact between the organic solvent and the virus. For many of the nonionic materials useful herein, the wetting agent can comprise from 0.001% to 10%, preferably from 0.01 to 2%, of the aqueous mixture, depending upon the amount of fatty material in the treated aqueous mixture. The amounts of solvent and detergent will also vary depending upon each other, upon the aqueous mixture being treated and the polypeptide to be purified.

The prior art has taught the addition of organic solvents and detergents to concentrated solutions of polypeptides to disrupt and inactivate lipid-enveloped viruses while preserving the protein structure of the desired polypeptide. See U.S. Patent No. 4,540,573. Organic solvents containing either Tween-80 or Triton X-100 have been shown to be effective reagents for killing viruses found in concentrated solutions of certain proteins without adversely affecting the bioactivity of the proteins. The use of such solvent-detergent mixtures, however, has been avoided because subsequent removal of the mixture from the concentrated protein has proven to be very difficult. See, for example, Prince, A.M. et al., *Lancet*, p. 706, March 29, 1981. As a result, other detergents, such as sodium cholate or sodium deoxycholate, are more commonly used. These detergents can be removed from the polypeptide of interest by gel exclusion chromatography on, for example, Sephadex G-25. A drawback to the use of these detergents, however, is that they are strong protein denaturants which can adversely affect the bioactivity of the polypeptide. Such detergents can be used herein in amounts which will not denature the polypeptide to be purified, either in combination with other detergents or with protein stabilizers or both.

The disadvantages of the methods of the prior art methods have been overcome by the preferred embodiments of the present invention. The present invention provides a means for the effective removal of virus-inactivating reagents from the polypeptide of interest, thus overcoming the obstacle to the use of detergents such as Tween 80 and Triton X-100 with an organic solvent. In the method of the present invention, the virus-inactivating agents are added to the complex aqueous material containing the polypeptide of interest. As described in greater detail below, the mixture then can be directly applied to a matrix which contains active immobilized monoclonal antibodies to the polypeptide. The polypeptide is adsorbed by the matrix, and the virus-killing reagents can be removed by extensive washing of the matrix.

The matrix can be an immunoaffinity chromatography column which contains a solid support matrix to which active monoclonal antibodies, able to specifically adsorb the Factor VIII:C, are coupled. The monoclonal antibody can be added to the resin support, which frequently has been activated with an activator such as cyanogen bromide. The support is made in accordance with conventional procedures and may comprise, for example, beaded agarose, cellulose, nylon or polyvinyl membranes. As mentioned above, the immunoaffinity matrix is usually in the form of a chromatographic column or cartridge. The preparation of such columns and cartridges is well within the skill of the art. Such columns or cartridges containing an appropriate support can be flushed with water and then equilibrated with the same buffer solution as the aqueous mixture containing the polypeptide to be purified.

The column then is washed with an aqueous buffer to remove a major portion of the virus-inactivating agent and other contaminants which may be non-specifically bound to the monoclonal antibody matrix or to the polypeptide without eluting the polypeptide adsorbed to the column.

The prior art, Livingston, D.M., *Methods in Enzymology*, 34, 723-731, (1974), has suggested that the non-specific binding of contaminants to the polypeptide of interest or the solid support matrix by ionic interaction can be minimized by using high ionic strength solutions or detergents. This has not proved to be a practical suggestion, for the addition of high ionic strength solutions and/or detergents to a conventional matrix often causes the polypeptides bound to the matrix to become unstable or causes a poor rate of

adsorption of the polypeptide to the matrix.

In the present process, however, high ionic strength polypeptide solutions containing detergent have been shown not to produce any adverse effects during the immuno-adsorption step when using monoclonal antibodies with hydrophobic attraction for the polypeptide. For example, the Factor VIII:C polypeptide from several source material solutions with high ionic strength, e.g. 0.5 to 1.5 M NaCl, were found to adsorb to an immunoaffinity matrix at a faster rate than with polypeptide solutions with low ionic strength, e.g. 0.15 M NaCl. Factor VIII:C is stable in such source solutions containing an organic solvent and detergent as well. When the complex aqueous mixture containing the polypeptide of interest and the virus-inactivating agents is added to the column, the polypeptide is adsorbed by the column and its constituents. Other elements of the mixture pass through the column.

After the immunoaffinity chromatography column has been sufficiently washed with the buffer that a major portion of the virus-inactivating agents and other contaminants have been removed from the polypeptide bound to the column, the column is treated with an agent which will release the bound polypeptide. The agent desirably is a polarity-reducing buffer with low ionic strength.

It has been found that using a low polarity, low ionic strength buffer to elute the polypeptide from the immunoaffinity column is advantageous, for the resulting polypeptide solution can be added without modifications to an ion exchange or other affinity matrix.

A major concern in conventional immunoaffinity purification has been the leaching of non-human monoclonal antibodies into the polypeptide eluate solution during the elution step. In the prior art, it is a common practice to change the pH, polarity, or ionic strength of the eluted polypeptide solution so that proper adsorption of the polypeptide to a second affinity region matrix can take place. Usually this alteration is sufficient to cause the leached monoclonal antibodies either to reassociate with the polypeptide or to bind to the second affinity matrix along with the polypeptide and thereby contaminate the final product. In the present process, however, the composition of the solution which desorbs the polypeptide from the immunoaffinity column does not need to be altered prior to the loading of the ion-exchange column. It therefore is more difficult for any monoclonal antibodies which may be present in the solution to rebind to either the polypeptide of interest or the second affinity matrix.

When the low polarity buffer solution is passed through the affinity matrix, the polypeptide binds to the affinity matrix and the solution, while any leached monoclonal antibodies, detergents or other contaminants pass through the column. The affinity matrix is preferably an ion-exchange column which comprises a matrix covalently coupled with charged chemical groups that are able to bind charged biological molecules under low ionic strength and release them as the pH or ionic strength changes. Suitable affinity materials include those chemical functional groups having no substantial affinity for antibodies. Examples of ionic chemical species include aminoalkyl anion exchangers, especially those where the alkyl groups have up to six carbon atoms and cation exchangers. Quaternary aminoethyl, aminoethyl and diethylaminoethyl compounds constitute one preferred class of anion-exchanging moieties. Sulfopropyl and carboxymethyl are acceptable and preferred cation-exchanging moieties. The affinity adsorbent support can be any of the commonly used supports such as fibers, beads, discs or platelets of crosslinked cellulosic material, beaded agarose and the like.

The matrix is then washed to remove remaining traces of contaminating residues on the polypeptide. The washing should be sufficient to reduce contaminants to a level non-toxic to humans, as well as to a level where they do not affect the efficacy of the polypeptide being purified. The washing solution preferably contains the same components as the eluting solution, but at a higher pH and/or lower ionic strength if an anion-exchange matrix with positively charged groups is used. As a general guideline, the elution solution should be buffered with a composition to provide a sufficiently high ionic strength and/or low pH to desorb the polypeptide from an anionic exchange matrix. The solution must be compatible with the polypeptide, be non-toxic, and be able to disrupt the polypeptide affinity matrix bond.

The solution containing the eluted polypeptide can be directly diluted into a buffered solution to produce a purified product suitable for therapeutic use. The solution typically comprises physiological concentrations of salts and human albumin along with nontoxic reagents such as amino acids, acetate, phosphate and citrate, imidazole, trimethamine and the like, that provide a buffering capacity of about pH 7.

The source material for the polypeptide to be purified includes plasma or plasma-derived fractions, for example, cryoprecipitate, suitably solubilized, which contain the polypeptide to be purified. Human plasma or plasma from other animals may also be used in this invention. Other source materials useful in this invention include human or animal tissue culture media containing a polypeptide produced by recombinant DNA technology.

To use cryoprecipitate in this invention, the cryoprecipitate can be dissolved by suspension in an aqueous solution containing components required to provide a reasonably stable environment for polypep-

5 tide activity. Such solution components include salts to maintain a suitable ionic strength. After suspension and thorough mixing of the cryoprecipitate in the aqueous solution, it is often desirable to remove insoluble material by such physical techniques as centrifugation or filtration. Adjustments to the cryoprecipitate solution, for example, pH, salts, and PEG addition, can also be made to facilitate the removal of insoluble material. The result is a particle-free solution with stable polypeptide activity. Other source materials may not require adjustments.

10 A second step can involve the addition of organic solvents and detergents to the source material solution to cause inactivation of lipid-enveloped viruses. After mixing of the organic solvent/detergent mixture with source material, the mixture and source material should stand for a sufficient period of time to inactivate lipid-enveloped viruses. The period of time is dependent on the temperature of the source material solution. A minimum of 3 minutes is used when the source material is a cryoprecipitate solution containing 0.3% tri-n-butyl phosphate and 1.0% Triton X-100 at a temperature of 18° C or greater.

15 Prior to performing the immunoaffinity purification step, it is desirable to optimize the solution conditions to enhance adsorption of the polypeptide by the immunoaffinity matrix. Such conditions include the addition of salts (e.g. sodium chloride, calcium chloride) in sufficient concentration to disrupt the association of polypeptide with other proteins if the latter are present. Such conditions could also include a pH adjustment to disrupt the association of polypeptide with other proteins. In general, a wide variety of physical and chemical techniques can be employed to optimize polypeptide adsorption by the immunoaffinity matrix. The techniques of choice will be obvious to one experienced in the field once the properties of the antibody preparation and immunoaffinity matrix have been established.

20 The immunoaffinity purification step in the process can be performed in a variety of ways. The objects of this step are (1) to adsorb polypeptide onto a matrix to which antibody has been first covalently bound, (2) to wash nonspecifically bound material from the immunoaffinity matrix, thereby separating contaminants from the polypeptide to be purified, and (3) to elute the polypeptide from the immunoaffinity matrix in a substantially purified form. The polypeptide-recognition sites on the antibody molecules are preferably always equal to or in excess of the polypeptide concentration so that quantitative adsorption of the polypeptide from the source material solution is possible. When the immunoaffinity matrix is contained in a column, the rate at which the source material is applied is adjusted to allow sufficient time for polypeptide adsorption to occur. When the immunoaffinity matrix is not contained in a column but is instead added to the polypeptide source material, as in a batch reaction, the mixture is stirred for a sufficient time to allow polypeptide adsorption to occur. It is a common practice to determine empirically the amount of time required for adsorption to occur when either reaction mode is used.

25 The polypeptide is adsorbed to the immunoaffinity matrix, and the immunoaffinity matrix is washed with an aqueous solution to remove nonspecifically bound or retained material from the matrix. Nonspecifically bound or retained material includes such components of the polypeptide source material as proteins, phospholipids, salts, organic solvent, detergent, and pyrogens or virus particles if the latter are present. The composition of the aqueous wash solution is such as to cause the removal of the above-mentioned nonspecifically bound or retained material from the immunoaffinity matrix, to retain polypeptide on the immunoaffinity matrix, and to maintain polypeptide activity. The composition of the aqueous wash solution meeting the above criteria is commonly determined empirically by those experienced in the field as is the amount of aqueous wash solution required to cause the substantial removal of nonspecifically bound or retained material from the immunoaffinity matrix. Since the polypeptide is Factor VIII:C, the ionic strength of the wash solution can have a major effect on the degree of purification achieved. This is because a high ionic strength wash solution removes Factor VIII:RP (von Willebrand Factor) from the immunoadsorbed Factor VIII:C. When followed by elution, the dissociation will result in a Factor VIII:C solution high in specific activity.

30 Following the washing of the immunoaffinity matrix, the polypeptide is eluted with an aqueous solution containing components causing the dissociation of polypeptide from the antibody. The components in the aqueous solution include polar or nonpolar materials which disrupt the noncovalent bonds that otherwise maintain the polypeptide bound to the antibody. The aqueous solution may contain other components which serve to maintain polypeptide activity, for example, calcium chloride and albumin. The components of the aqueous solution and their respective concentrations can be determined once the other parameters of polypeptide type and complex aqueous mixture type have been selected.

35 A second part of the inventive process is the use of an affinity matrix purification. Some of the most notable benefits of the process are realized by the two-step process where the immunoaffinity purification precedes the affinity purification. After the polypeptide mixture has been added to the affinity matrix, the anionic exchange affinity region is washed with a low-ionic strength buffer solution. The region is then eluted with a buffer salt solution which preferably has a high ionic strength and/or low pH, and components

which serve to maintain polypeptide stability, e.g. calcium ions, polyethylene glycol, and albumin. The eluate containing the polypeptide of interest can be collected in a solution which is the equivalent of the eluting solution except that the eluting salt is deleted.

The process of the present invention is further illustrated by the following examples, which are provided for illustrative purposes only and are not to be construed as limiting.

The following is a list of the formulations for solutions employed in Examples 1 to 4.

Solution I (Immunoaffinity Equilibration Solution): 0.05 M imidazole, 0.8 M sodium chloride, 0.05 M calcium chloride, 0.3% (v/v) tri (n-butyl) phosphate, 1.0% Triton X-100; pH adjusted to 7.4 ± 0.1 with 6 N HCl.

Solution II (Immunoaffinity Wash Solution): 0.05 M imidazole, 0.04 M calcium chloride, 5.0% (v/v) ethylene glycol; pH adjusted to 6.4 ± 0.1 with 6 N HCl.

Solution III (Immunoaffinity Elution Solution): 0.05 M imidazole, 0.04 M calcium chloride, 40% (v/v) ethylene glycol, 0.1% (1.0 mg/ml) human albumin; pH adjusted to 6.4 with 6 N HCl.

Solution IV (QAE-ZetaPrep Wash Solution): 0.05 M histidine, 0.15 M sodium chloride, 1.0 mM calcium chloride, 1.0 M glycine, 0.1% (w/v) polyethylene glycol 4000, 0.1% human albumin (U.S.P.); pH adjusted to 6.4 with 6 N HCl.

Solution V (QAE-ZetaPrep Elution Solution): 0.05 M histidine, 0.6 ± 0.1 M sodium chloride, 4.0 mM calcium chloride, 0.1% (w/v) polyethylene glycol 4000, 1% human albumin (U.S.P.); pH adjusted to 5.5 to 6.4 with 6 N HCl.

Solution VI (Bulk Dilution Solution): 0.05 M histidine, 4.0 mM calcium chloride, 0.1% (w/v) polyethylene glycol 4000, 1.0% human albumin (U.S.P.); pH adjusted to 8.2 with 6 N HCl.

Solution VII : 0.05 M histidine, 0.15 M sodium chloride, 4.0 mM calcium chloride, 0.1% (w/v) polyethylene glycol 4000, 1.0% human albumin (U.S.P.), pH adjusted to 7.1 ± 0.1 with 6 N HCl.

EXAMPLE 1

A. Preparation of the Anti-Factor VIII:C Monoclonal Antibody Solution

The monoclonal antibody used herein is derived from a hybridoma, which hybridoma was prepared following generally the method of Milstein and Köhler. Following immunization of Balb/c female mice with human Factor VIII:C, the spleen cells of the immunized mice were fused with myeloma cells P-3Ag 8653 from a mouse myeloma and the resultant hybridomas were screened for those supernatants containing antibody which give selective binding to human Factor VIII:C. The desired hybridoma was subsequently cloned and characterized. As a result, a hybridoma with the identification number G1-F8/1.5.6, from Genetics Institute, Cambridge, Massachusetts, was obtained which produces antibody against an epitope on the human Factor VIII:C protein. Not only does this antibody react with human Factor VIII:C obtained from plasma, but it has been found that it also reacts with human Factor VIII:C produced by cells transformed with recombinant DNA coding for human Factor VIII:C. The preparation of the monoclonal antibody used herein was carried out as follows:

Immunization

Female Balb/c mice (obtained from Jackson Labs) are immunized intraperitoneally on day 0 with 25-50 units of Factor VIII obtained as set forth in Example I which had been emulsified in 0.4ml of complete Freund's adjuvant. On day 21, the mice are reimmunized with the Factor VIII emulsified in incomplete Freund's adjuvant. Subsequent booster immunizations are administered at three week intervals.

The sera of the immunized animals are tested three days after each boost for inhibition of Factor VIII coagulant activity by incubating dilutions of the sera with normal pooled plasma at 37°C for 2 hours. The residual Factor VIII activity is then measured by the chromogenic Factor VIII assay.

Three days prior to fusion, the mice are challenged with a final injection (intravenous or intraperitoneal) of 25-50 units of Factor VIII in phosphate buffered saline.

Cell Fusion

Fusion is carried out in the absence of serum in accordance with the procedure developed by Köhler and Milstein. The spleens from two immunized mice are removed by blunt dissection, cut longitudinally, followed by teasing out the cells on a plate into 2 mls of ice-cold RPMI-1640/Glutamine, plate rinsed once with 2 ml of the same buffer to get residual cells, and placed in 10 ml tubes. The debris is allowed to settle

out for about 5 minutes on ice and the cells are washed at 4°C in 50 mls RPMI-1640/Glutamine and thereafter suspended in 10 mls of ice-cold RPMI-1640/Glutamine. A count of the cells totaled 2×10^8 .

Myeloma cells of strain P3Ag8653 maintained in 207 FCS/RPMI (glutamine, 2-mercapto ethanol, Gentamycin) are washed at 10°C in 50 ml RPMI-1640/Glutamine and thereafter suspended in 10 ml of room temperature RPMI-1640.

The spleen and myeloma cells are mixed in a ratio of about 4 spleen cells to every myeloma cell and brought up to 50 ml RPMI-1640/Glutamine and washed at 10°C. The supernatant is aspirated and the pellet resuspended by flicking the tube. The tube is then placed in a beaker containing 37°C water. One ml of 50% w/w PEG 1500 is added gradually to the tube over 1/2 minute while stirring the pellet with the tip of a 1 ml pipette. The mixture is allowed to stand 1 1/2 minutes at 37°C with occasional stirring. Five ml of RPMI-1640/Glutamine at 37°C is gradually added over a three minute period while stirring. An additional 14 ml of RPMI-1640/Glutamine is added over 1 minute. Followed by 30 ml of 20% FCS RPMI-1640/Glutamine. The cell suspension is centrifugated at 20°C for 8 minutes and the pellet is suspended to 1.5×10^5 /ml (1.5×10^7 spleen cells/plate/20 ml) input spleen cells/ml in 20% FCS RPMS-1640 + HAT. Normal mouse peritoneal cell feeders (2.5×10^4 /ml) from the same strain as the spleen cell donor is added. 0.2 ml aliquots were placed drop wise in Costar 96 well places (measure out 20 ml/plate). After 7 days 1/2 - 2/3 of the medium is replaced with HAT + RPMI-1640 + 20% FCS. Between days 12-28 the medium is replaced periodically with HT & RPMI-1640 & 20% FCS (the medium is replaced when the cells began to turn yellow: -2 days). From day 30 on, the medium is replaced with RPMI-1640 + 20% FCS.

The stock solutions are as follows:

1. HT 200X: hypoxanthine, 136 mg
thymidine, 38.8 mg
D.D.W., 100 ml
dissolved at 70°C, filter sterilized, and stored frozen at -29°C for about 6 months.
2. Aminopterin 1,000X: aminopterin 3.5 mg
0.10 N NaOH, 1 ml
D.D.W., 19 ml
filter sterilized, stored in dark at -20°C for -2 months.
3. HAT 1X: HT 100X, 5 ml; Aminopterin 1.000X, 0.5 ml; RPMI + 20% FCS, 500 ml
4. HT 1X: HT 100X, 0.5 ml; RPMI + 20% FCS, 500 ml
5. Azaguanine 100x: 8-azaguanine 2.5 mg
0.01 N NaOH 10 ml
filter sterilized, stored in 1 ml aliquots at 20°C

Screening Hybrid Cell Cultures for Antibodies to Factor VIII:C

Hybrid cell cultures are screened for the production of anti-Factor VIII:C antibodies by two methods.

1. Binding assay

Microtiter plates (96-well) are coated with rabbit anti-mouse IgG by incubating in 0.2 M carbonate buffer, pH 9.5 for two hours at about 37°C. The plates are washed three times with phosphate buffered saline containing 0.05% Tween 20 (PBS/Tween 20). Nonspecific sites were coated by incubating the plates with PBS containing 3% gelatin. Aliquots of hybrid cell supernatant are added to the wells and incubated at 37°C for two hours and thereafter washed three times with PBS/Tween 20. A plasma-derived fraction of human Factor VIII:C is added and incubated overnight at room temperature and thereafter washed three times with PBS/Tween 20. Anti-Factor VIII 125 I-FAB' (prepared from IgG isolated from the plasma of a patient with a high titer Factor VIII inhibitor), is added and incubated at 37°C for four hours and subsequently washed three times in PBS. The wells are cut with a hot wire cutter and a radioactivity count is made. Positives in this assay indicated antibody to Factor VIII:C.

2. Inhibition Assay

Aliquots of hybrid cell supernatant are mixed with equal volumes of normal pooled plasma and incubated at 37°C for 2 hours. The samples are diluted to obtain a proper assay (chromogenic or clotting assay) of Factor VIII:C bioactivity. A decrease in the Factor VIII:C activity of normal pooled plasma indicates the presence of an inhibitory antibody to Factor VIII:C. This assay does not detect the presence of non-inhibitory antibodies to Factor VIII:C.

Cells which are positive by either of the above two assays are cloned and subcloned in soft agar in accordance with the technique described herein below. Subcloned cells are then grown in mice as ascites tumors for the production of antibody-rich ascites fluid.

5 Soft Agar Cloning Technique

1. Prepare 0.5% agar medium:
For 500 ml:
10X Earl's Balanced Salt Solution, 5.5 ml, Mix and warm to 45 °C.
10 RPMI-1640, 434 ml
Heat 50 ml of 5% agar solution to 100 °C,
and add the heated solution to the RPMI-1640 solution.
Incubate in 45 °C waterbath.
Allow for 18 ml 0.5% agar/plate.
- 15 2. Hybrid cell dilutions: for each line use 4 tubes labeled with line's name and marked 1, 8⁻¹, 8⁻², 8⁻³.
Dilutions may vary, e.g., 8⁻¹, 8⁻², 8⁻³, 8⁻⁴. To tube 1 add 0 ml, to others 0.7 ml of RPMI-1640. Hybrid
cells should be healthy and in the log phase of growth. The cells are suspended and 0.7 ml transferred
to tube 1 and 0.1 to tube 8⁻¹, mixed and 8-fold serial dilutions are carried out by diluting 0.1 ml from the
tube 8⁻¹ into tubes 8⁻², 8⁻³ and discarding 0.1 from 8⁻³ tube, so all contain 0.7 ml. Place in 37 °C bath.
20 It is preferred that all of these dilutions are used for first cloning. The '1' dilution may usually be omitted
for subcloning.
3. 100 x 15 mm petri dishes are labeled and with a 25 ml disposable pipette 15 ml of 0.5% Bacto-agar
medium is pipetted into dishes and allowed to harden for 20 minutes before adding the soft agar overlay
containing cells in 0.33% agar medium. It is important not to tilt plates during this period. The remainder
25 of the 0.5% Bacto-agar medium is kept at 45 °C.
4. Soft agar overlay: An aliquot of 0.5% agar is poured into a 50 ml plastic tube and placed in a 45 °C
water bath in the hood. Tubes containing cells are removed individually from the 37 °C bath. 1.4 ml agar
is added with a 5 or 10 ml disposable pipette, pipetted up and down to mix, and most of the mixture
(avoiding bubbles) is overlayed onto the 0.5 agar base. It is essential during this step that neither the
30 agar nor cells have cooled, otherwise a homogeneous 0.33% agar mixture will not be obtained.
5. The plates are allowed to harden for 30 minutes without moving (except for sliding in hood) and
incubated at 37 °C - 5% CO₂ for 7-14 days.
6. When discrete colonies appear, clones are 'picked' with sterile pasteur pipettes (i.e., the tip is placed
over the colony and sucked up using a clean pipette for each colony), and the cells are placed in 150 ul
35 of 20% FCS/RPMI-1640 in 96 well Costar plates.
7. The plates are incubated at 37 °C 5% CO₂ for 7-14 days. When culture supernatant turns yellow, the
clones are screened by the defined methods.

Purification of Monoclonal IgG from Ascites Fluid

- 40 A. Ascites fluid from Example IV is filtered through glass wool plug to remove particulate matter.
 - B. The Ascites fluid from (A) is passed through 0.45 micron filter, diluted with an equal volume of 0.02 M
Tris, pH 8.5 and passed through 0.22 micron filter immediately prior to purification as below.
 - 45 C. The filtered sample is injected onto a TSK DEAE-SPW HPLC ion exchange column equilibrated at
24 °C in 0.02 M Tris, pH 8.5. The flow rate is adjusted to 1.0 ml per minute, and 1-ml fractions are
collected. Bound IgG is eluted using a linear gradient from 0% B to 30% B (buffer A is 0.02 M Tris, pH
8.5; buffer B is 0.02 M Tris, pH 7.0 containing 1.0 M NaCl).
 - D. Fractions are assayed for inhibition of Factor VIII:C bioactivity by mixing an aliquot of each fraction
with appropriately diluted normal pooled plasma prior to analysis in a Factor VIII:C bioassay (as
50 described above).
 - E. Fractions containing anti-VIII:C antibody are pooled for use as immuno-adsorbent.
- The hybridoma, prepared as described above, was procured from Genetics Institute, Inc. of Cambridge,
Massachusetts.
- Monoclonal antibodies (mAb) to human plasma Factor VIII:C are dialyzed against ten volumes of 0.1 M
55 sodium bicarbonate, 0.5 M NaCl, pH 8.5 (coupling solution) for 4 hours at 2 to 8 °C. The dialysis is repeated
overnight. The antibody protein concentration, determined by adsorption at 280 nm, is adjusted to 1.1
grams per liter with the coupling solution.

B. Preparation of the Immunoaffinity Resin

The monoclonal antibody is coupled to cyanogen bromide-activated Sepharose CL-2B as described by March et al, *Anal Biochem*, 60, 149-152 (1974). The method is as follows. Sepharose CL-2B, pre-swollen, is washed with ten resin-volumes of deionized water, and suspended in one resin-volume of cold (2-8°C) deionized water. Two resin-volumes of 2.0 M sodium carbonate are added to the resin and stirred. For each liter of resin, 60 g of cyanogen bromide is dissolved in 120 ml of acetonitrile and added to the resin with vigorous stirring. After 10 to 30 minutes of incubation, depending upon the rate and extent of activation needed, the resin suspension is suction-filtered and washed five times with two resin-volumes of coupling solution. The resin is immediately transferred to a beaker and added to the dialyzed monoclonal antibody solution. The resin suspension is stirred for two hours at 21°C which produces a coupling efficiency greater than 90% with approximately 1 g of monoclonal antibody bound per liter of resin. The resin suspension is stored overnight in the cold, suction filtered, and washed four times with two resin-volumes of cold coupling solution. The resin is transferred to a beaker containing three resin-volumes of 0.2 M glycine, pH 8.0, stirred for two hours at 21°C to block the remaining reactive sites on the resin, suction filtered, and washed five times with two resin-volumes of a 0.1 M sodium acetate, 0.5 M NaCl, pH 3.5 solution. The antibody-resin mixture is finally washed four times with two resin-volumes of 10 mM acetic acid, pH 3.5, and stored in one resin-volume of the same solution at 2-8°C until needed. The antibody-resin mixture has been found to be functionally active when stored over prolonged periods in the acetic acid solution.

C. Preparation of Factor VIII:C Source Material Solution

Approximately 3000 liters of fresh human plasma is mixed with an anticoagulant, and stored at -25°C. The frozen human plasma is thawed under conditions which result in the formation of 30 kg of insoluble material referred to as cryoprecipitate (cryo) as described by Shanbrom et al. U.S. Patent 3,631,018 (1971). The cryo is collected by centrifugation which separates the cryo from the soluble plasma material. The cryo, whether it had been stored frozen at -70°C or received fresh at 5°C, is dissolved at 20 to 30°C in two volumes of distilled water containing 50 µM CaCl₂. The pH of the cryo suspension is slowly adjusted to 6.7 ± 0.2 by the addition of 1 M acetic acid. The temperature is lowered to 9°C and held at that temperature until an insoluble material forms over time and is separated from the soluble material by centrifugation. The temperature of the cold supernate (about 75 liters) is increased to 21°C and the pH adjusted to 7.4 ± 0.2 with 1 N NaOH. Sodium chloride (solid) and calcium chloride (5 M stock) are slowly added to the cold supernate solution to obtain final concentrations of 0.8 M NaCl and 0.05 M CaCl₂, respectively. Finally, a 3.33 to 1 ratio mixture of Triton X-100 and tri(n-butyl) phosphate (TNBP) is added to the solution as a viral inactivating agent at a final concentration of 1.3% (v/v). The solution thus obtained is referred to as cryoprecipitate-detergent (cryo-detergent) solution.

D. The Immunoaffinity Chromatography Step

Approximately 2.5 liters of the antibody-resin mixture is packed into a column in the presence of 10 mM acetic acid, pH 3.5 at 21°C. The column is washed at 21°C with 3 to 4 resin-volumes of distilled water followed by equilibration with 3 to 4 resin-volumes of Solution I. Approximately 75 liters of cryo-detergent solution is applied to the column from top to bottom at a flow rate of 2 to 5 liter per hour. The column is washed with 18 resin-volumes of Solution II at a flow rate of 10 liter per hour. The Factor VIII:C polypeptide is eluted off the column in the reverse flow direction by applying about four resin-volumes of Solution III. The flow rate ranges from 2 to 5 liters per hour.

E. The Ion Exchange Chromatography Step

The ion exchange matrix, which is a compressed cellulose disk cartridge containing quaternary aminoethyl functional groups (QAE ZetaPrep 250) is conditioned for use by washing the cartridge with the following solutions: 500 ml of 0.15 M NaCl, 1 liter of 1.0 M acetic acid, 1 liter of 0.1 M imidazole, 1.0 M NaCl, pH 8.2, eight liters of 0.15 M NaCl, and finally two liters of Solution III. After conditioning, the mAb eluate solution containing the Factor VIII:C is loaded onto the QAE ZetaPrep 250 at a flow rate ranging from 0.4 to 1.0 liter per hour. The QAE ZetaPrep cartridge is washed with two liters of Solution IV and Factor VIII:C is eluted with about two liters of Solution V. The eluate is added to 3 volumes of physiologically compatible Solution VI, and further diluted with Solution VII to adjust the final potency of the Factor VIII:C. The ultrapurified Factor VIII:C material is sterile filtered through a 0.2 micron filter, placed in glass vials,

frozen, and lyophilized without heat treatment.

EXAMPLE 2

5 A. Preparation of Factor VIII:C Source Material Solution

Approximately 86 kg of cryoprecipitate derived from plasma as described in Example 1, is stored at -70 °C as a human Factor VIII:C enriched plasma fraction, is dissolved in twice its weight of water and calcium chloride, is added to a final concentration of 50 uM. The pH of the cryosuspension (250 liters) is slowly adjusted to 6.7 with 1 M acetic acid. The temperature of the Factor VIII:C solution is gradually cooled to 9.5 °C as a precipitation occurs over a 20-minute period of continuous mixing. The precipitate, which is predominately fibrinogen and fibronectin accounting for about 50% of the total protein, is removed by centrifugation at 3000 x g for 15 minutes at 9.5 °C. The pH of the resulting cold supernate (225 liters) is adjusted to 7.4 with 1 N NaOH, and filtered through a 0.5 to 0.8 micron normal pore size membrane to remove any particles with little loss of Factor VIII:C. To the filtered supernate is added NaCl to 0.8 M, CaCl₂ to 0.05 M, and a Triton X-100/tri-(n-butyl) phosphate mixture (3.3:1 ratio) to 1.3% (v/v) with little loss of Factor VIII:C (17.9 units/ml). This is designated as the cryo-detergent solution.

Immunoaffinity Chromatography Step

The cryo-detergent solution, 230 liters, is applied at room temperature from top to bottom through a vertical column containing 5 liters of the immunoabsorbent (capacity is 1 g mAb per liter resin) at a flow rate of 8.5 liters per hour. The mAb-resin is washed with 40 volumes of Solution II at a flow rate of 20 liters per hour to remove contaminating proteins, viruses, and the virus-inactivating agents.

The resin-bound Factor VIII:C is desorbed from the resin with 19.4 liters of Solution III at a flow rate of 6 liters per hour.

Ion-Exchange Chromatography Step

The Factor VIII:C mAb eluate pool (19.4 liters) is passed directly onto a QAE-ZetaPrep 800 cartridge (LKB, Bromma, Sweden) from the inner core through to the outer peripheral matrix at a flow rate of 1 to 5 liters per hour. The QAE-ZetaPrep matrix is washed with 8 to 10 liters of Solution IV at a flow rate of 4 liters per hour. The Factor VIII:C is eluted from the QAE matrix in a reverse flow direction with 3 liters of Solution V at a flow rate of 3.2 liters per hour, and added to 9 liters of Solution VI. Finally, the potency of Factor VIII:C bulk solution is adjusted with 6.5 liters of Solution VII to produce a highly purified Factor VIII:C suitable for therapeutic use.

EXAMPLE 3

This example demonstrates that increasing the ionic strength of the Factor VIII preparation will increase the rate of Factor VIII:C adsorption to resin coupled with anti-Factor VIII:C monoclonal antibodies which bind to Factor VIII:C through hydrophobic interaction. Results are presented in Table 1.

In the first experiment, cryoprecipitate was dissolved in a solution such that it comprised 9.5 units/ml Factor VIII:C in a solution of 0.03 M sodium citrate, 0.12 M NaCl, 0.1 M glycine, and 1 unit/ml heparin. One ml aliquots of the cryoprecipitate solution, containing Factor VIII:C in the presence of 0.12, 0.52, and 1.0 M NaCl, respectively, were each added to 0.1 ml monoclonal antibody-resin and 0.1 ml control resin coupled with normal mouse IgG. With continuous mixing, 0.1 ml aliquots of the resin slurries were centrifuged and assayed for Factor VIII:C activity at time t=0, 0.5, 2.5, and 5 hours.

In the second experiment, Hemofil C (antihemophilic factor, Baxter-Hyland Division) was reconstituted with water such that it comprised 28 units/ml Factor VIII:C in a solution of 0.02 M sodium citrate, 0.15 M NaCl, 0.10 glycine, 50 uM CaCl₂, 1% (w/v) polyethylene glycol, 1% human albumin, pH 7.0. Fourteen ml aliquots of Factor VIII:C solutions containing 0.15 M, and 1.0 M NaCl, respectively, were each added to 1 ml mAb-resin and 1 ml uncoupled control resin. With continuous mixing, 0.3 ml aliquots of the resin slurries were centrifuged and assayed for Factor VIII:C activity at time t=0, 0.08, 0.25, 0.5, 1.0, 1.5, and 2.0 hours.

In the third experiment, lyophilized mAb-purified Factor VIII:C was reconstituted with water such that it comprised 14 units/ml Factor VIII:C in Solution VII. The rate of Factor VIII:C adsorption to the mAb-resin was determined as described in the second experiment of this example.

TABLE 1

| IONIC STRENGTH EFFECT ON THE RATE OF FACTOR VIII:C ADSORPTION TO mAb-RESIN | | | |
|--|---|-------------------------|--|
| Experiment | # Factor VIII Preparations | Final Conc. of NaCl (M) | Relative Factor VIII:C Activity Adsorbed after 1 hr. |
| 1 | Cryo Solution Cryo Solution Cryo Solution | 0.12 0.52 1.0 | 27 43 53 |
| 2 | Hemofil C, AHF Hemofil C, AHF | 0.15 1.0 | 54 62 |
| 3 | Method M, AHF* Method M, AHF* | 0.15 1.0 | 89 97 |

* Monoclonal antibody-purified Factor VIII:C

EXAMPLE 4

This example demonstrates that the eluted Factor VIII:C material from the antibody-resin composite is highly purified, although retaining a significant portion of Factor VIII:RP associated with it.

A quantity of 1.15 kg of cryoprecipitate was dissolved in twice its weight in water in the presence of 50 μ M CaCl_2 . The pH was adjusted to 6.7 with acetic acid and gradually cooled down to 9.5°C. The precipitation which formed over a 10 minute period was removed by centrifugation at 4,500 xg for 20 min. The pH of the cold temperature supernate was adjusted to 7.4 with 1 M NaOH followed by addition of NaCl to 0.8 M, CaCl_2 to 0.05 M, and a 3.3:1 ratio of Triton X-100: tri-(n-butyl) phosphate mixture to 1.3% (v/v). The solution is referred to below as cryo-detergent solution.

Approximately 3 liters of the cryo-detergent solution was applied at room temperature from top to bottom through a 100 ml column of mAb-resin (capacity: 1 mg mAb per ml resin) at a flow rate of 3 ml/min. The column was washed with 1800 ml of Solution II at a low rate of 10 ml/min. The Factor VIII:C material was eluted off the mAb resin with 450 ml of Solution III without human albumin at a low rate of 5 ml/min. The specific activity of the purified Factor VIII:C was then determined by one-stage clotting assays, and protein concentration derived from absorbance at 280 nm and dye-binding techniques. Table 2 below shows the activity of the Factor VIII:C recovered from the indicated starting material.

TABLE 2
PURIFICATION OF FACTOR VIII:C BY THE IMMUNOAFFINITY METHOD

| STEP | TOTAL ACTIVITY | VOLUME | PROTEIN | SPECIFIC ACTIVITY (Units/mg) | PURIFI- CATION | RECOVERY |
|-----------------|--|---------|---------|------------------------------------|-------------------|----------|
| | (Units) | (ml) | (g) | | | (%) |
| Human Plasma | 100,000 | 120,000 | 7,000 | 0.014 | 1 | 100 |
| Cryoprecipitate | 53,800 | 3,450 | 180 | 0.30 | 21 | 54 |
| Suspension | | | | | | |
| Cryo-detergent | 52,000 | 3,100 | 84 | 0.62 ^a | 44 | 52 |
| Solution | | | | | | |
| Immunoaffinity | 43,500 | 450 | 0.024 | 1800 ^b | 128,000 | 44 |
| Eluate | | | | | | |
| a | contains approximately 0.17 Units Factor VIII:RP per Unit Factor VIII:C | | | | | |
| b | contains approximately 0.010 Units Factor VIII:RP per Unit Factor VIII:C | | | | | |

Claims

1. A method for purifying Factor VIII:C from a mixture of polypeptides and other constituents comprising:
 - (a) immobilizing an antibody, which antibody binds by hydrophobic attraction to the Factor VIII:C to be purified, to a matrix before or after said Factor VIII:C is added to said antibody, thereby immobilizing said Factor VIII:C in an immunoaffinity matrix;
 - (b) eluting the Factor VIII:C from the immobilized antibody by treating the Factor VIII:C:immunoaffinity matrix with a desorbing substance to desorb the Factor VIII:C from said matrix, thereby forming a Factor VIII:C:desorbing substance mixture;
 - (c) passing the Factor VIII:C to be purified through a second region capable of binding to the Factor VIII:C through a hydrophilic attraction, thereby binding the Factor VIII:C to the second region while allowing contaminants to pass through the second region;
 - (d) wherein the Factor VIII:C:desorbing substance mixture is passed from the matrix of step (b) to the region of step (c) without further modification of the said mixture; and
 - (e) eluting the purified Factor VIII:C from the said second region.
2. A method according to Claim 1 wherein said mixture of polypeptides is blood plasma.
3. A method according to Claim 1 wherein said mixture of polypeptides is a blood plasma fraction that has been solubilized.
4. A method according to Claim 1 wherein said mixture of polypeptides is present in a cell culture medium derived from recombinant DNA technology.
5. A method according to Claim 1 wherein said mixture of polypeptides is present in a final product which is to be repurified.

6. A method according to any one of the preceding claims wherein said immunoaffinity matrix contains an antibody coupled to a matrix.
7. A method according to Claim 6 wherein said antibody is monoclonal.
8. A method according to Claim 7 wherein said monoclonal antibody is specific for Factor VIII:C.
9. A method according to Claim 6 wherein said matrix comprises a resin or agarose beads.
10. A method according to any one of the preceding claims wherein a washing is employed after the Factor VIII:C to be purified is adsorbed by the immunoaffinity matrix.
11. A method according to Claim 10 wherein said washing is carried out with a buffered salt solution.
12. A method according to any one of the preceding claims wherein said desorbing substance is a buffered salt solution containing a nonpolar agent.
13. A method according to Claim 12 wherein said nonpolar substance is ethylene glycol or propylene glycol.
14. A method according to any one of the preceding claims wherein said immunoaffinity matrix is a vertical column, a radial-flow cartridge, or a free mixture of said matrix with polypeptide.
15. A method according to Claim 1 wherein said second region contains insoluble particles, resin, agarose beads or cellulose.
16. A method according to Claim 19 wherein said second region is an ion exchange matrix containing amino diethylaminoethyl or quaternary aminoethyl functional groups as anion exchangers, or sulfopropyl, phospho or carboxyalkyl functional groups as cation exchangers.
17. A method according to Claim 16 wherein said matrix contains an amino diethylaminoethyl or quaternary aminoethyl functional group deposited on a cellulosic matrix in a cartridge.
18. A method according to any one of the preceding claims wherein a washing is employed after the polypeptide is bound to the second region.
19. A method according to Claim 18 wherein said washing is conducted with a buffered salt solution of low ionic strength.
20. A method according to any one of the preceding claims wherein the eluting of the Factor VIII:C from either the immunoaffinity matrix or the second region matrix is accomplished by adding the eluting solution to said matrix to cause a flow through the matrix in a reverse direction from the flow of the Factor VIII:C mixture or Factor VIII:C solution into the matrix.
21. A method according to any one of the preceding claims wherein a virus-inactivating stage is employed.
22. A method according to Claim 21 wherein said virus-inactivating stage uses an organic solvent and/or a detergent to inactivate the viruses present in the mixture of polypeptides.
23. A method according to Claim 22 wherein said organic solvent is a tri-n-alkyl phosphate, dialkyl ether or amyl acetate.
24. A method according to Claim 22 wherein said detergent is Triton X-100, Tween 80, sodium cholate or sodium deoxycholate.
25. A method according to Claim 22 wherein said organic solvent is a tri-n-butyl phosphate and said detergent is an oxyethylated alkylphenol.

26. A method according to any one of Claims 21 to 25 wherein said virus-inactivating stage is performed prior to the immunoaffinity purification stage or step (c).
27. A process for purifying Factor VIII:C from a complex aqueous mixture contaminated with lipid-enveloped pathogenic viruses, which comprises:
- (a) mixing the complex aqueous mixture with a virus-inactivating agent containing an organic lipid-dissolving or disrupting solvent and a detergent at any stage in the process;
 - (b) passing the complex aqueous mixture through an immunoaffinity matrix containing active immobilized monoclonal antibody specific to the Factor VIII:C and which binds by hydrophobic attraction to said Factor VIII:C, thereby causing the Factor VIII:C to adsorb to the immunoaffinity matrix;
 - (c) washing the immunoaffinity matrix with an aqueous buffer to remove at least a portion of contaminants from the complex aqueous mixture, while leaving the Factor VIII:C adsorbed to the immunoaffinity matrix;
 - (d) eluting the Factor VIII:C from the immunoaffinity matrix with a desorbing substance, thereby forming a Factor VIII:C:desorbing substance mixture;
 - (e) passing the Factor VIII:C:desorbing substance mixture through a second region, thereby causing the Factor VIII:C to bind thereto, while allowing any leached monoclonal antibodies, detergents and other contaminants to pass through the second region, wherein the Factor VIII:C:desorbing substance mixture is passed from the matrix of step (c) to the matrix of step (e) without further modification of the said mixture; and
 - (f) eluting the second region with an eluting solution to desorb the Factor VIII:C to form a solution substantially free of contaminants.
28. A method according to Claim 27 wherein said organic lipid-disrupting solvent in step (a) is tri-(n-butyl) phosphate, and said detergent is Triton X-100.
29. A method according to Claim 27 wherein said complex aqueous mixture in step (b) has a high ionic strength comprising sodium chloride ranging from 0.5 to 1.5 molar or calcium chloride ranging from 0.07 to 0.5 molar, and said monoclonal antibody has a hydrophobic attraction for Factor VIII:C.
30. A method according to Claim 27 wherein said aqueous buffer in the step (c) is a low ionic strength buffered salt solution.
31. A method according to Claim 27 wherein said eluting low polarity buffer in step (d) is a low ionic strength buffered salt solution containing ethylene glycol or propylene glycol, and has a pH ranging from 6.3 to 7.2.
32. A method according to Claim 27 wherein said second region in step (e) contains amino diethylaminoethyl or quaternary aminoethyl functional groups deposited on a cellulosic matrix in a cartridge.
33. A method according to Claim 27 wherein said eluting solution for the affinity matrix in step (f) is a high ionic strength buffered salt solution containing sodium chloride ranging from 0.6 to 0.8 molar, human albumin ranging from 0.1 to 1 per cent, and has a pH ranging from 5.5 to 6.4.

Patentansprüche

1. Verfahren zur Reinigung von Faktor VIII:C aus einem Gemisch von Polypeptiden und anderen Bestandteilen umfassend:
- (a) Immobilisierung eines Antikörpers, welcher sich durch hydrophobe Anziehung an den zu reinigenden Antikörper bindet, an einer Matrix bevor oder nachdem der Faktor VIII:C dem Antikörper zugesetzt wird, wodurch der Faktor VIII:C an einer Immunoaffinitätsmatrix immobilisiert wird;
 - (b) Eluieren des Faktors VIII:C vom immobilisierten Antikörper, indem die Faktor VIII:C Immunoaffinitätsmatrix zur Desorption von Faktor VIII:C von der Matrix mit einer desorbierenden Substanz behandelt wurde, wodurch ein Faktor VIII:C desorbierendes Substanzgemisch gebildet wird;
 - (c) Durchleiten des zu reinigenden Faktor VIII:C durch einen zweiten Bereich, der fähig ist, den Faktor VIII:C durch hydrophile Anziehung zu binden; dadurch wird ein Bindung des Faktor VIII:C an

- den zweiten Bereich erreicht, während Verunreinigungen den zweiten Bereich passieren gelassen werden;
- (d) wobei das Faktor VIII:C: desorbierende Substanzgemisch von der Matrix von Stufe (b) zu dem Bereich von Stufe (c) ohne eine weitere Veränderung des Gemisches geführt wird; und
- 5 (e) Eluieren des gereinigten Faktor VIII:C aus dem zweiten Bereich.
2. Verfahren nach Anspruch 1, bei dem das Gemisch aus Polypeptiden Blutplasma ist.
 3. Verfahren nach Anspruch 1, bei dem das Gemisch aus Polypeptiden eine Blutplasmafraktion ist, die
10 solubilisiert wurde.
 4. Verfahren nach Anspruch 1, bei dem das Gemisch aus Polypeptiden in einem Zellkulturmedium vorliegt, das aus der Gentechnologie abgeleitet ist.
 - 15 5. Verfahren nach Anspruch 1, bei dem das Gemisch aus Polypeptiden in einem Endprodukt vorliegt, das wieder zu reinigen ist.
 6. Verfahren nach einem der vorangehenden Ansprüche, bei dem die Immunoaffinitätsmatrix einen an eine Matrix gebundenen Antikörper enthält.
20
 7. Verfahren nach Anspruch 6, bei dem der Antikörper monoklonal ist.
 8. Verfahren nach Anspruch 7, bei dem der monoklonale Antikörper für Faktor VIII:C spezifisch ist.
 - 25 9. Verfahren nach Anspruch 6, bei dem die Matrix ein Harz oder Agarose-Kügelchen enthält.
 10. Verfahren nach einem der vorangehenden Ansprüche, bei dem ein Waschen vorgenommen wird, nachdem der zu reinigende Faktor VIII:C durch die Immunoaffinitätsmatrix adsorbiert worden ist.
 - 30 11. Verfahren nach Anspruch 10, bei dem das Waschen mit einer gepufferten Salzlösung durchgeführt wird.
 12. Verfahren nach einem der vorangehenden Ansprüche, bei dem die desorbierende Substanz eine gepufferte Salzlösung ist, die ein nichtpolares Agens enthält.
 - 35 13. Verfahren nach Anspruch 12, bei dem die nichtpolare Substanz Ethylenglykol oder Propylenglykol ist.
 14. Verfahren nach einem der vorangehenden Ansprüche, bei dem die Immunoaffinitätsmatrix eine senkrechte Säule, eine Radialflußhülle oder eine freie Mischung aus Matrix und Polypeptid ist.
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 15. Verfahren nach Anspruch 1, bei dem der zweite Bereich unlösliche Teilchen, Harz, Agarose-Kügelchen oder Cellulose enthält.
 - 45 16. Verfahren nach Anspruch 19, bei dem der zweite Bereich eine Ionenaustauschermatrix ist, die als Anionenaustauschergruppen Aminodiethylaminoethyl- oder quarternäre Aminoethylgruppen oder als Kationenaustauschergruppen Phospho- oder Carboxyalkylgruppen enthält.
 17. Verfahren nach Anspruch 16, bei dem die Matrix eine Aminodiethylaminoethyl- oder quarternäre Aminoethylgruppe als funktionelle Gruppe enthält, die an einer Cellulosematrix in einer Hülle angelagert ist.
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 18. Verfahren nach einem der vorangehenden Ansprüche, bei dem ein Waschvorgang zwischengeschaltet wird, nachdem das Polypeptid an den zweiten Bereich gebunden worden ist.
 - 55 19. Verfahren nach Anspruch 18, bei dem der Waschvorgang mit gepufferter Salzlösung einer niedrigen Ionenkonzentration durchgeführt wird.

20. Verfahren nach einem der vorangehenden Ansprüche, bei dem die Elution von Faktor VIII:C entweder von der Immunoaffinitätsmatrix oder von der Matrix des zweiten Bereichs erreicht wird, indem die eluierende Lösung der Matrix zugesetzt wird, um eine Strömung in entgegengesetzter Richtung zu der Strömung der Faktor VIII:C-Mischung oder der Faktor VIII:C-Lösung in der Matrix zu erzeugen.
21. Verfahren nach einem der vorangehenden Ansprüche, bei dem eine virusinaktivierende Stufe eingebaut ist.
22. Verfahren nach Anspruch 21, bei dem die virusinaktivierende Stufe ein organisches Lösungsmittel und/oder ein oberflächenaktives Mittel zur Inaktivierung der im Polypeptidgemisch vorliegenden Viren verwendet.
23. Verfahren nach Anspruch 22, bei dem das organische Lösungsmittel ein Tri-n-alkylphosphat, Dialkylether oder Amylacetat ist.
24. Verfahren nach Anspruch 22, bei dem das oberflächenaktive Mittel Triton X-100, Tween 80, Natriumcholat oder Natriumdeoxycholat ist.
25. Verfahren nach Anspruch 22, bei dem das organische Lösungsmittel ein Tri-n-butylphosphat und das oberflächenaktive Mittel ein oxyethyliertes Alkylphenol ist.
26. Verfahren nach einem der Ansprüche 21 bis 25, bei dem die virusinaktivierende Stufe vor der Immunoaffinitäts-Reinigungsstufe oder Stufe (c) durchgeführt wird.
27. Verfahren zur Reinigung von Faktor VIII:C aus einer komplexen wässrigen Mischung, die mit lipidumhüllten pathogenen Viren kontaminiert ist, umfassend:
 - (a) Vermischen der komplexen wässrigen Mischung mit einem virusinaktivierenden Agens, das ein organisches lipidlösendes oder -spaltendes Lösungsmittel und ein oberflächenaktives Mittel enthält, in irgendeiner Stufe im Verfahren;
 - (b) Hindurchleiten der komplexen wässrigen Mischung durch eine Immunoaffinitätsmatrix, die einen aktiven immobilisierten monoklonalen Antikörper enthält, der spezifisch für Faktor VIII:C ist und der sich durch hydrophobe Anziehung an den Faktor VIII:C bindet, wodurch bewirkt wird, daß Faktor VIII:C an der Immunoaffinitätsmatrix adsorbiert wird;
 - (c) Waschen der Immunoaffinitätsmatrix mit einem wässrigen Puffer, um mindestens einen Teil der Verunreinigungen aus der komplexen wässrigen Mischung zu entfernen, während der Faktor VIII:C an der Immunoaffinitätsmatrix adsorbiert bleibt;
 - (d) Eluieren des Faktor VIII:C von der Immunoaffinitätsmatrix mit einer desorbierenden Substanz, dadurch Bildung einer Faktor VIII:C: desorbierende Substanz-Mischung;
 - (e) Durchleiten der Faktor VIII:C: desorbierenden Substanz-Mischung durch einen zweiten Bereich; wodurch bewirkt wird, daß Faktor VIII:C daran gebunden wird, während durchgesickerte monoklonale Antikörper, oberflächenaktive Mittel und andere Verunreinigungen den zweiten Bereich passieren gelassen werden, wobei die Faktor VIII:C: desorbierende Substanz-Mischung die Matrix von Stufe (c) bis Matrix von Stufe (e) ohne weitere Veränderung der Mischung passiert; und
 - (f) Eluieren des zweiten Bereichs mit einer eluierenden Lösung, um den Faktor VIII:C unter Bildung einer im wesentlichen von Verunreinigungen freien Lösung zu desorbieren.
28. Verfahren nach Anspruch 27, bei dem das organische lipidspaltende Lösungsmittel in Stufe (a) Tri (n-butyl)-phosphat und das oberflächenaktive Mittel Triton X-100 ist.
29. Verfahren nach Anspruch 27, bei dem die komplexe wässrige Mischung in Stufe (b) eine hohe Ionenkonzentration hat und Natriumchlorid in einer 0,5 bis 1,5-molaren Konzentration oder Calciumchlorid in einer 0,07 bis 0,5-molaren Konzentration enthält, und bei dem der monoklonale Antibody eine hydrophobe Anziehung für Faktor VIII:C hat.
30. Verfahren nach Anspruch 27, bei dem der wässrige Puffer in Stufe (c) eine gepufferte Salzlösung mit niedriger Ionenkonzentration ist.

31. Verfahren nach Anspruch 27, bei dem der eluierende Puffer mit niedriger Polarität in Stufe (d) eine gepufferte Salzlösung mit niedriger Ionenkonzentration ist, die Ethylenglykol oder Propylenglykol enthält und einen pH-Wert im Bereich von 6,3 bis 7,2 hat.
- 5 32. Verfahren nach Anspruch 27, bei dem der zweite Bereich in Stufe (e) Aminodiethylaminoethyl- oder quarternäre Aminoethyl-Gruppen als funktionelle Gruppen enthält, die an einer Cellulosematrix in einer Hülse angelagert sind.
33. Verfahren nach Anspruch 27, bei dem die eluierende Lösung für die Affinitätsmatrix in Stufe (f) eine gepufferte Salzlösung mit niedriger Ionenkonzentration ist, die Natriumchlorid in 0,6 bis 0,8-molarer Konzentration, menschliches Albumin in 0,1 - 1%-Konzentration enthält, und die einen pH-Wert im Bereich von 5,5 bis 6,4 hat.
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Revendications

- 15 1. Méthode de purification du Facteur VIII:C à partir d'un mélange de polypeptides et autres composants comprenant:
- (a) l'immobilisation d'un anticorps, ledit anticorps étant lié par attraction hydrophobe au Facteur VIII:C à purifier, à une matrice avant ou après addition dudit Facteur VIII:C audit anticorps, immobilisant ainsi ledit Facteur VIII:C dans une matrice d'immunoaffinité;
- 20 (b) l'élution du Facteur VIII:C à partir de l'anticorps immobilisé en traitant la matrice d'immunoaffinité du Facteur VIII:C avec une substance éluante pour désorber le Facteur VIII:C de ladite matrice, formant ainsi un mélange de substance désorbant le Facteur VIII:C;
- (c) le passage du Facteur VIII:C à purifier à travers une deuxième zone capable de se lier au Facteur VIII:C par attraction hydrophile, liant ainsi le Facteur VIII:C à la deuxième zone tout en permettant le passage des contaminants à travers la deuxième zone;
- 25 (d) le mélange de substance désorbant le Facteur VIII:C étant passé de la matrice de la phase (b) à la zone de la phase (c) sans autre modification dudit mélange; et
- (e) l'élution du Facteur VIII:C purifié de ladite seconde zone.
- 30 2. Méthode selon la revendication 1 dans laquelle ledit mélange de polypeptides est du plasma sanguin.
3. Méthode selon la revendication 1 dans laquelle ledit mélange de polypeptides est une fraction de plasma sanguin qui a été solubilisée.
- 35 4. Méthode selon la revendication 1 dans laquelle ledit mélange de polypeptides se trouve dans un milieu de culture cellulaire dérivé par technologie d'ADN recombinant.
5. Méthode selon la revendication 1 dans laquelle ledit mélange de polypeptides se trouve dans un produit fini à repurifier.
- 40 6. Méthode selon l'une quelconque des revendications précédentes dans laquelle ladite matrice d'immunoaffinité contient un anticorps couplé à une matrice.
7. Méthode selon la revendication 6 dans laquelle ledit anticorps est monoclonal.
8. Méthode selon la revendication 7 dans laquelle ledit anticorps monoclonal est spécifique pour le Facteur VIII:C.
- 50 9. Méthode selon la revendication 6 dans laquelle ladite matrice contient une résine ou des perles d'agarose.
10. Méthode selon l'une quelconque des revendications précédentes dans laquelle un lavage est effectué après que le Facteur VIII:C à purifier a été adsorbé par la matrice d'immunoaffinité.
- 55 11. Méthode selon la revendication 10 dans laquelle ledit lavage est effectué avec une solution saline tamponnée.

12. Méthode selon l'une quelconque des revendications précédentes dans laquelle ladite substance désorbante est une solution saline tamponnée contenant un agent non polaire.
13. Méthode selon la revendication 12 dans laquelle ladite substance non polaire est de l'éthylène glycol
ou du propylène glycol.
14. Méthode selon l'une quelconque des revendications précédentes dans laquelle ladite matrice d'immunoaffinité est une colonne verticale, une cartouche à flux radial ou un mélange libre de ladite matrice avec un polypeptide.
15. Méthode selon la revendication 1 dans laquelle ladite seconde zone contient des particules insolubles, de la résine, des perles d'agarose ou de la cellulose.
16. Méthode selon la revendication 19 dans laquelle ladite seconde zone est une matrice échangeuse d'ion contenant des groupes fonctionnels amino diéthylaminoéthyle ou aminoéthyle quaternaire comme échangeurs d'anions, ou des groupes fonctionnels sulfopropyle, phosphoalkyle ou carboxyalkyle comme échangeurs de cations.
17. Méthode selon la revendication 16 dans laquelle ladite matrice contient un groupe fonctionnel amino diéthylaminoéthyle ou aminoéthyle quaternaire déposé sur une matrice cellulosique dans une cartouche.
18. Méthode selon l'une quelconque des revendications précédentes dans laquelle un lavage est effectué après que le polypeptide est lié à la seconde zone.
19. Méthode selon la revendication 18 dans laquelle ledit lavage est effectué avec une solution saline tamponnée de faible force ionique.
20. Méthode selon l'une quelconque des revendications précédentes dans laquelle l'élution du Facteur VIII:C à partir de la matrice d'immunoaffinité ou de la matrice de la seconde zone est effectuée en ajoutant la solution éluante à la dite matrice afin de provoquer un flux à travers la matrice dans une direction inverse de celle du flux du mélange de Facteur VIII:C ou de la solution de Facteur VIII:C dans la matrice.
21. Méthode selon l'une quelconque des revendications précédentes dans laquelle une phase d'inactivation virale est utilisée.
22. Méthode selon la revendication 21 dans laquelle ladite phase d'inactivation virale utilise un solvant organique et/ou un détergent pour inactiver les virus présents dans le mélange de polypeptides.
23. Méthode selon la revendication 22 dans laquelle ledit solvant organique est du tri-(n-alkyl) phosphate, un éther dialkyle ou de l'acétate d'amylo.
24. Méthode selon la revendication 22 dans laquelle ledit détergent est du Triton X-100, du Tween 80, du cholate de sodium ou du déoxycholate de sodium.
25. Méthode selon la revendication 22 dans laquelle ledit solvant organique est du tri-(n-butyl) phosphate et ledit détergent un alkylphénol oxyéthylé.
26. Méthode selon l'une quelconque des revendications 21 à 25 dans laquelle ladite phase d'inactivation virale est effectuée avant la phase de purification par immunoaffinité ou l'étape (c).
27. Procédé de purification du Facteur VIII:C à partir d'un mélange aqueux complexe contaminé par des virus pathogènes à enveloppe lipidique, qui comprend :
(a) le mélange du mélange aqueux complexe avec un agent d'inactivation virale contenant un solvant organique dissolvant les lipides ou disruptif des lipides et un détergent à toute étape du procédé;

- (b) le passage du mélange aqueux complexe à travers une matrice d'immunoaffinité contenant l'anticorps monoclonal immobilisé actif spécifique pour le Facteur VIII:C et qui se lie par attraction hydrophobe audit Facteur VIII:C, le Facteur VIII:C s'adsorbant alors à la matrice d'immunoaffinité;
- 5 (c) le lavage de la matrice d'immunoaffinité avec un tampon aqueux pour éliminer au moins une partie des contaminants du mélange aqueux complexe, tout en laissant le Facteur VIII:C adsorbé à la matrice d'immunoaffinité;
- (d) l'élution du Facteur VIII:C à partir de la matrice d'immunoaffinité avec une substance désorbante, formant ainsi un mélange de substance désorbant le Facteur VIII:C.
- 10 (e) le passage du mélange de substance désorbant le Facteur VIII:C à travers une deuxième zone, entraînant la liaison du Facteur VIII:C à cette zone tout en laissant passer les anticorps monoclonaux lessivés, les détergents et les autres contaminants à travers la seconde zone, le mélange de substance désorbant le Facteur VIII:C passant de la matrice de l'étape (c) à la matrice de l'étape (e) sans autre modification dudit mélange; et
- 15 (f) l'élution de la seconde zone avec une solution éluante pour désorber le Facteur VIII:C pour former une solution sensiblement exempte de contaminants.
28. Méthode selon la revendication 27 dans laquelle ledit solvant organique disruptif des lipides de l'étape (a) est le tri-(n-butyl) phosphate et ledit détergent le Triton X-100.
- 20 29. Méthode selon la revendication 27 dans laquelle ledit mélange aqueux complexe de l'étape (b) a une force ionique élevée comprenant du chlorure de sodium compris entre 0,5 et 1,5 mol ou du chlorure de calcium compris entre 0,07 et 0,5 mol et ledit anticorps monoclonal ayant une attraction hydrophobe pour le Facteur VIII:C.
- 25 30. Méthode selon la revendication 27 dans laquelle ledit tampon aqueux de l'étape (c) est une solution saline tamponnée de faible force ionique.
31. Méthode selon la revendication 27 dans laquelle ledit tampon éluant de faible polarité de l'étape (d) est une solution saline tamponnée de faible force ionique contenant de l'éthylène glycol ou du propylène glycol, avec un pH compris entre 6,3 et 7,2.
- 30 32. Méthode selon la revendication 27 dans laquelle la seconde zone de l'étape (e) contient des groupes fonctionnels amino diéthylamnioéthyle ou aminoéthyle quaternaire déposés sur une matrice cellulosique dans une cartouche.
- 35 33. Méthode selon la revendication 27 dans laquelle ladite solution éluante pour la matrice d'affinité de l'étape (f) est une solution saline tamponnée de force ionique élevée contenant du chlorure de sodium, compris entre 0,6 et 0,8 mol, de l'albumine humaine comprise entre 0,1 et 1 % et possède un pH compris entre 5,5 et 6,4.
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